Testosterone measurements in early infancy

C Tomlinson, H Macintyre, C A Dorrian, S F Ahmed, A M Wallace


Although "direct" commercial steroid immunoassays are reliable in many clinical situations, concerns remain about their accuracy for steroid measurements in young infants. Determination of the plasma concentration of testosterone is an important part of the evaluation of young infants with genital abnormalities.

The results confirm that neonatal circulating testosterone concentrations are method dependent, and each laboratory should establish method related reference ranges especially if using a direct commercial immunoassay. The results indicate that the Bayer ADVIA Centaur procedure can be used reliably in neonates. Expected values for male and female infants < 10 days old were 2.5-11.1 (n = 36) and 1.7-5.6 (n = 36) nmol/l respectively. For older neonates (10-50 days) the ranges were 0.2-17.2 (n = 42) and 0.1-1.5 (n = 7) nmol/l respectively.

RESULTS

Circulating testosterone concentrations in infants measured by a direct chemiluminescent immunoassay (Bayer ADVIA Centaur) were compared with those measured by a traditional radioimmunoassay using solvent extraction. The results confirm that neonatal circulating testosterone concentrations are method dependent, and each laboratory should establish method related reference ranges especially if using a direct commercial immunoassay. The results indicate that the Bayer ADVIA Centaur procedure can be used reliably in neonates. Expected values for male and female infants < 10 days old were 2.5-11.1 (n = 36) and 1.7-5.6 (n = 36) nmol/l respectively. For older neonates (10-50 days) the ranges were 0.2-17.2 (n = 42) and 0.1-1.5 (n = 7) nmol/l respectively.

DISCUSSION

Concern existed in the literature that commercial assays were inaccurate in the clinical setting of newborn infants when there are high levels of sulphated steroids. In fact, a spuriously raised testosterone concentration has occurred in our laboratory in a neonatal sample using the Bayer Immuno-1 system, leading to diagnostic confusion. Fortunately, no spuriously raised results were obtained by the direct method used in this study. Although results from the two assays correlated well, there were significant differences in the values obtained. For neonates under 10 days of age, significantly lower testosterone concentrations were obtained by the in house extraction assay. This could be related to removal of cross reacting conjugated steroids by solvent extraction. For the older boys, significantly lower results were obtained by the direct assay, which is more difficult to explain. It is, however, clear that the direct assay was specifically developed for measuring testosterone in adults. The testosterone protein binding characteristics in neonatal samples are considerably different from that in adults, and this may affect the accuracy of the measurement. Further studies are required to clarify this issue.

Our results confirm that neonatal circulating testosterone concentrations are method dependent and each laboratory should establish method related reference ranges especially if using a direct commercial immunoassay. In addition to the ADVIA Centaur analyser used in this study, there are at least eight commercial non-radioactive immunoassays for testosterone. Each of these methods has been developed and optimised for measurement of testosterone in adults and should be viewed as suspect in relation to neonatal testosterone measurements unless proven otherwise.

Using the extracted assay, we studied circulating testosterone concentrations in detail in both sexes over the first 6 months of life. As previously reported in boys, the concentrations are in general very high at delivery showing a very wide variation, with almost a 10-fold difference in girls less than 10 days old (n = 36 and 36 respectively) and between 10 and 50 days old (n = 42 and 7 respectively). The overall correlation between assays was good (r = 0.88; n = 121). In both boys and girls aged less than 10 days, significantly lower results (p < 0.001) were obtained by the extraction radioimmunoassay. In boys > 10 days, significantly lower results were obtained by the direct procedure (p < 0.001).

Figure 1 shows the plasma testosterone concentrations measured by the extraction assay in boys and girls. In boys plasma testosterone concentrations are high at birth but rapidly fall over the first week before rising to a peak at 2 months of age. In girls, testosterone concentrations remain generally low over the first 6 months but could be as high as 5 nmol/l at birth.

SUBJECTS AND METHODS

Blood samples were obtained from a group of infants less than 6 months old having other investigations. Ethical approval was obtained from Yorkhill NHS Trust. Testosterone was measured directly by chemiluminescent immunoassay on the Bayer ADVIA Centaur analyser (sensitivity 0.35 nmol/l, coefficient of variation < 7.6%) and an in house radioimmunoassay with a preceding solvent extraction step (sensitivity 0.35 nmol/l, coefficient of variation < 8%).

Intergroup comparison was performed by the paired t test using SPSS software.

RESULTS

Table 1 shows the median concentration and range of plasma testosterone as assayed by both the Centaur ADVIA direct immunoassay and the extraction radioimmunoassay in boys and girls less than 10 days old (n = 36 and 36 respectively) and between 10 and 50 days old (n = 42 and 7 respectively). The overall correlation between assays was good (r = 0.88; n = 121). In both boys and girls aged less than 10 days, significantly lower results (p < 0.001) were obtained by the extraction radioimmunoassay. In boys > 10 days, significantly lower results were obtained by the direct procedure (p < 0.001).
between some infants. This concentration then falls very rapidly before a slow rise to a second peak at 2 months and then falling to "normal" prepubertal concentrations by 6 months of age. Again there is a very wide variation, with a 10-fold difference between some infants. In girls, there is a much smaller peak at delivery, which then falls to low concentrations over the first week, which remain low over the next six months.

In conclusion, we have shown that there are small differences in measured testosterone concentrations in infancy between a commercial direct assay and one using an extraction step. These differences are, however, unlikely to cause problems in a clinical setting, and it would appear that the widely available Bayer ADVIA Centaur direct assay can be used reliably to measure testosterone in plasma from newborn infants.

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**REFERENCES**

**Table 1** Median and range of matched samples for testosterone assays by sex and age

<table>
<thead>
<tr>
<th></th>
<th>In house extraction assay</th>
<th>Bayer ADVIA Centaur direct assay</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>n</td>
<td>Median Range</td>
</tr>
<tr>
<td><strong>Boys</strong> &lt;10 days</td>
<td>36</td>
<td>2.3 1.0–11.5*</td>
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<tr>
<td>&gt;10 days</td>
<td>42</td>
<td>4.6 0.3–29.3*</td>
</tr>
<tr>
<td><strong>Girls</strong> &lt;10 days</td>
<td>36</td>
<td>0.7 0.3–3.6*</td>
</tr>
<tr>
<td>&gt;10 days</td>
<td>7</td>
<td>0.5 0.2–1.3**</td>
</tr>
</tbody>
</table>

Values are nmol/l.
*Reference range (within 95% confidence intervals).
**Absolute range (insufficient data to calculate 95% confidence intervals).