Glucose-6-phosphate dehydrogenase activity in male premature and term neonates

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Background: Glucose-6-phosphate dehydrogenase (G6PD) deficiency is a common defect, affecting hundreds of millions of people, with a worldwide distribution.1 Its incidence varies from < 3% in the United States and Europe to 25% in some parts of Africa and the Middle East. Within any given area, the incidence may vary between population subgroups.2 In Israel, in some subsets of Sephardic Jews there is a high incidence of the G6PD Mediterranean mutation.3 Although best known for acute, life-threatening haemolytic crises (favism), G6PD deficiency is also associated with severe neonatal jaundice, with the potential of bilirubin encephalopathy, or kernicterus, if untreated.4–6 In a recent report of the informal United States based Kernicterus Registry, at least 21% of readmitted infants with kernicterus had documented G6PD deficiency.7 Premature neonates are susceptible to kernicterus at lower concentrations of serum total bilirubin than are term neonates. It is therefore possible that in population groups with a high incidence of G6PD deficiency, this condition may play a role in the pathogenesis of kernicterus in premature infants. However, there are few studies of G6PD deficiency in premature infants.

Normal neonates born at term have been shown to have higher G6PD activity than adults,8 and some studies have suggested even higher activity in premature infants.9–11 We therefore hypothesised that premature infants would have even higher G6PD activity than term neonates, and asked whether this increase would influence the ability to diagnose the deficiency state. In this study, we determined, quantitatively, normal values for red blood cell G6PD activity in male premature neonates and compared these values with those of healthy term and near term male infants.

METHODS
Clinical protocol
The study was approved by the Institutional Review Boards of both Bnai Zion and Shaare Zedek Medical Centers and further sanctioned by the Israeli Government Ministry of Health. The amount of blood required for study purposes was small and was drawn simultaneously with blood sampling for routine laboratory tests. In this study there was no randomisation of patients, clinical or therapeutic trial, additional risk to the babies or any other deviation from routine clinical management. For these reasons, the authorities gave approval to perform the study without the need for individual parental consent.

Any male, preterm infant, ≤ 36 weeks gestation, admitted to the premature care nursery of the participating centres, was eligible for the study. Neonates with major congenital anomalies, documented sepsis, intrauterine growth retardation, or grade 3 or 4 intraventricular haemorrhage apparent at the time of the G6PD testing were excluded. Neonates with respiratory distress or receiving ventilator treatment were not excluded from the study, as such a restriction would have resulted in the exclusion of almost every neonate ≤ 29 weeks gestation and many of those with higher gestational ages as well. Furthermore, we are unaware of any data suggesting that mechanical ventilation for respiratory distress syndrome influences G6PD activity. Gestational age, for the purpose of the study, was defined as the number of completed weeks subsequent to the first day of the last menstrual period, validated by ultrasound examinations during the pregnancy and confirmed by physical examination after delivery. In the event of a discrepancy, or if the gestational age could not be accurately determined, that baby was excluded from analysis. In addition to the premature infants, 24 healthy term and near term male neonates (gestational ages ≥ 37 weeks) were sampled for comparison. The study was limited to male infants, as G6PD deficiency is an X-linked condition and, in males, who can be either normal or deficient hemizygotes, the G6PD status can be accurately determined. In females, on the other hand, a high number of

Abbreviations: G6PD, glucose-6-phosphate dehydrogenase; Hb, haemoglobin
G6PD deficient heterozygotes, a form of the condition that may be difficult to diagnose, may be encountered.\(^1\)

Within 48 hours of delivery, and before any blood had been administered by transfusion, 0.25–0.5 ml blood was collected in an EDTA-containing microtainer. In those neonates who had an indwelling vascular catheter in place, this blood was collected through the catheter. In the remaining babies, blood was sampled by venepuncture, at the time of blood sampling for a routine laboratory investigation. No baby was hypoglycaemic, hypoxic, or had metabolic acidosis at the time of sampling. This blood was filtered through cellulose to remove leucocytes,\(^2\) and the filtrate stored at −70°C pending laboratory analysis.

**Laboratory methods**

Before the G6PD assay, the haemolysate was thawed and haemoglobin (Hb) concentration was measured by the Coulter method (Coulter T-980; Coulter Electronics, Luton, UK). G6PD activity was determined using a commercial kit (G6PD: quantitative, ultraviolet kinetic determination in blood at 340 nm, kit no 345 UV; Sigma Diagnostics, St Louis, Missouri, USA). The principle of the test involves the oxidation of glucose 6-phosphate to 6-phosphogluconate, and the concomitant reduction of NADP\(^+\) to NADPH. These reactions occur in the presence of G6PD, and the rate of NADPH formation, which is proportional to G6PD activity, is measured spectrophotometrically. One international unit (U) of activity is defined as the amount of G6PD activity that will convert 1 μmol glucose 6-phosphate to 6-phosphogluconate per minute.

A 0.01 ml portion of haemolysate was added to a vial containing G6PDH assay solution comprising NADP, maleimide, buffer, and stabilising agent, mixed, and left at room temperature for 5–10 minutes. G6PD substrate solution containing glucose 6-phosphate was added to the vial, and the contents transferred to a test cuvette. The cuvette was placed in a constant temperature water bath, and the absorbance at 340 nm read at baseline and five minutes later, versus potassium dichromate, in a narrow width spectrophotometer (Ocean Scientific Co, Garden Grove, California, USA). The rate of increase in absorbance at 340 nm was calculated and expressed as U/g Hb. The manufacturer’s reported normal range in healthy adults is 4.6–13.5 U/g Hb.

**Sample size**

Based on an estimated 25% increase in G6PD activity in pre-mature infants compared with term and near term neonates, with a power of 0.8 and an alpha value of 0.05, 17 neonates would be necessary in each group. The complete range of prematurity for study purposes (23–36 weeks) was divided arbitrarily into three approximately equal groups by gestational week, and at least 17 patients sampled in each group.

**Data analysis**

Based on a previous study from our laboratory in term infants, a cut off point of 5.0 U/g Hb enzyme activity was chosen to delineate the G6PD deficient from the G6PD normal infants.\(^13\) To facilitate determination of normal values for G6PD activity, neonates who were clearly G6PD deficient were not included in subsequent analysis. G6PD activity for the preterm infants (≤ 36 weeks gestation) were compared with those of the term and near term neonates (≥ 37 weeks gestation) using Student’s t test. When it had been confirmed that G6PD activity was higher in the premature neonates than in the term and near term group, the premature neonates were stratified into subgroups at two weekly intervals of gestational age, beginning at 23 weeks, and the subgroups compared using analysis of variance, to identify the sub-

![Figure 1](http://fn.bmj.com/Arch Dis Child Fetal Neonatal Ed) first published as 10.1136/adc.2004.049148 on 21 October 2004. Downloaded from http://fn.bmj.com/ on June 7, 2022 by guest. Protected by copyright.
neonates. There was no difficulty, however, in differentiating between G6PD normal and G6PD deficient neonates, as activity in the latter group was clearly lower than, and with no overlap between, that in the normal neonates.

Only a few studies of G6PD activity have been performed in premature or low birthweight neonates, and with inconsistent results. Marks and Gross\(^1^1\) compared premature and term newborns with adults, and found that G6PD activity in the former group was higher than in term infants of comparable age. However, their premature infant group included those aged 2 days to 1 year, and no data were available for gestational age or birth weight. Stewart and Birnbeck\(^13\) also studied premature infants, along with term neonates and adults. As in our study, they showed higher activity in premature than in term infants, and both these groups had higher activity than adults. Furthermore, decreasing gestational age correlated inversely with G6PD activity. However, in that series, no neonate of gestational age < 28 weeks or < 1000 g was included. Herz et al\(^1^4\) found higher G6PD activity in infants of low birth weight than normal counterparts. However, values for infants of birth weight 865–2160 g, with gestational ages of 28–40 weeks, were pooled, with no stratification for either birth weight or gestational age. In a group of relatively large premature infants (birth weight 2141 (310) g), Osiki et al\(^1^5\) documented that G6PD activity within the first 48 hours of life was significantly higher than in adults, but not significantly higher than in term infants.

In contrast with these studies, we prospectively studied G6PD activity in premature and term neonates enrolled shortly after birth, and found that activity in premature infants was higher than in term and near term neonates. The range of our study group extended from 23 weeks (the border of gestational viability) to 42 weeks gestation, and those born before 36 weeks stratified at two weekly intervals. Our results pinpointed the increase in G6PD activity in premature infants to the 29–32 week gestational period.

The peak G6PD activity at about 30 weeks gestation and subsequent decline in activity towards term is interesting and has not been previously described. The reason for the increase in activity is not clear. Had immaturity, or an immaturity related disease process, been a major factor in producing this phenomenon, the more immature babies would have been expected to have even higher G6PD activity than those at 29–32 weeks gestation. It was previously thought that the higher G6PD activity in newborns, compared to adults, was due, in part, to the large number of immature red blood cells and reticulocytes with higher G6PD activities, in this population subgroup. However, Konrad et al\(^1^6\) corrected for reticulocyte count, and showed that a high number of these cells was not a prerequisite for the increased G6PD activity in newborn blood. Mohrenweiser et al\(^1^7\) did find that newborns had higher G6PD activity than adults, although no significant differences were noted when blood with a high reticulocyte count was compared with samples with normal reticulocyte values. Additional as yet undetermined factors appear to be responsible for the intrinsically higher G6PD activity in neonates and premature infants. The peak we have described suggests a maturational effect contributing in part to the control of red blood cell G6PD activity in premature infants.