Background: Lung fibrosis is thought to be important in chronic lung disease of prematurity (CLD). Proliferative activity of lung lavage fluid (BALF) was studied in 207 bronchoalveolar lavage fluid (BALF) samples from 43 infants. Sixteen developed CLD (birth weight 765 g (630–1230), gestation 26.5 weeks (23–29)), 18 developed respiratory distress syndrome (RDS) (birth weight 1415 g (430–4160), gestation 31 weeks (23–39)), and nine control infants (birth weight 2110 g (900–3720), gestation 32 weeks (26–41)) received mechanical ventilation for non-pulmonary reasons.

Methods: The mitogenic activity relative to 10% fetal calf serum was 64–75% in infants with CLD, 55–86% in the RDS group, and 42–68% in control infants during the first 5 weeks of life. Only at day 3 there was a difference between the groups (CLD 72% v control 42%, p < 0.01; RDS 63% v control 42%, p < 0.05). With the use of neutralising antibodies, platelet derived growth factor BB (PDGF-BB) and epidermal growth factor were undetectable, and insulin-like growth factor I (IGF-I) accounted for 14% (p < 0.05) and 11% (p < 0.005) of BALF mitogenic activity from the RDS and CLD groups respectively.

Conclusions: The mitogenic activity of BALF was similar in the three groups studied and was only partially accounted for by IGF-I. Growth factors other than PDGF-BB and IGF-I contribute significantly to this process.

Although the risk factors for chronic lung disease of prematurity (CLD) have been accurately described, the underlying pathogenesis is less clear. Although oxygen toxicity and barotrauma are important risk factors, and a more recently described addition is pulmonary inflammation. Histologically, in infants who die from CLD, lung fibrosis is a characteristic feature of the disease. The hyaline membranes, which are present early in the development of neonatal respiratory distress syndrome (RDS), are replaced during the second week by interstitial and perivascular fibrosis. The septal walls are thickened, and myofibroblast proliferation is seen by electron microscopy. Using immunohistochemistry, we have shown that newly synthesised collagen, namely procollagen type I, is deposited early in infants who die from respiratory failure compared with those who die from non-respiratory causes. Components of the extracellular matrix, including N-termini of collagen molecules and fibronectin, are also increased in tracheal fluid obtained from infants who develop CLD. Thus pulmonary fibrosis appears to be a hallmark of CLD.

Development of pulmonary fibrosis is dependent on the proliferation of fibroblasts together with synthesis and deposition of the extracellular matrix by these cells. Proliferation of fibroblasts and the subsequent synthesis of the extracellular matrix is thought to be mediated by growth factors. Transforming growth factor β (TGF-β) is a potent profibrotic agent promoting the synthesis and deposition of the extracellular matrix. We have previously shown that it is considerably increased early in bronchoalveolar lavage fluid (BALF) obtained from infants who developed CLD compared with those infants who did not. The growth factors platelet derived growth factor B (PDGF-BB) and insulin-like growth factor I (IGF-I) are potent mitogens for fibroblasts.

IGF-I consists of B and A chains, with the mature 70 amino acid molecule being highly conserved in vertebrates, showing 40% homology with insulin and 60% with IGF-II. IGF-I is expressed in fetal tissues by mesenchymal cells, and its ablation in mice results in decreased prenatal growth and lethality immediately after birth. Postnatally, IGF-I has multiple functions, including acting as a progression factor for fibroblasts, thus inducing their proliferation. IGF-I has been shown to be increased in systemic sclerosis and in idiopathic pulmonary fibrosis. PDGF is a 30 kDa protein of either homodimer (PDGF-AA or PDGF-BB) or a heterodimer (PDGF-AB). Dimers with at least one B chain will bind to the receptor PDGFR-β. PDGF-BB is more relevant to respiratory biology as it is a potent mitogen for lung fibroblasts and has been shown to be increased in respiratory diseases such as idiopathic lung fibrosis and acute lung injury. Sources within the lung include alveolar macrophages, which are increased in many respiratory diseases including CLD.

As both IGF-I and PDGF-BB are potent mitogens for lung fibroblasts, and as lung fibrosis is a hallmark of CLD, we postulated that BALF obtained from infants who developed CLD would have greater fibroblast mitogenicity than similar fluid obtained from infants who required mechanical ventilation for neonatal RDS and for non-pulmonary reasons. In addition, by using antibody neutralising experiments, we determined whether PDGF-BB or IGF-I contributed to the fibroblast proliferative activity of BALF.

METHODS

Patient groups

Three groups of mechanically ventilated infants were studied: (a) CLD group: infants who developed RDS followed by CLD—that is, who were oxygen dependent at 28 days of age and had an abnormal chest radiograph; (b) RDS group:

Abbreviations: CLD, chronic lung disease of prematurity; RDS, respiratory distress syndrome; TGF, transforming growth factor; BAL, bronchoalveolar lavage; BALF, bronchoalveolar lavage fluid; PDGF-BB, platelet derived growth factor B; IGF-I, insulin-like growth factor I; FCS, fetal calf serum; FGM, fibroblast growth medium; EGF, epidermal growth factor.
infants who developed RDS but were nursed in air and had a normal chest radiograph by day 28 of age; (c) control group: infants who received mechanical ventilation for non-pulmonary reasons and had an oxygen requirement of ≤28% throughout the study period.

Bronchoalveolar lavage (BAL)

BAL was performed at the time of clinically indicated tracheal suctioning as previously described. Briefly, the infant was placed in the supine position with the head turned to the left to direct the catheter into the right lower lobe. An FG 5 catheter was gently inserted in a segmental bronchus until resistance was felt, and two aliquots of 1 ml/kg saline was instilled. With the catheter in situ, after two or three ventilator breaths, the instilled fluid was aspirated back with a suction pressure of 5–7 kPa. The procedure was performed at the time of clinically indicated routine suctioning. The first BAL was performed within 24 hours of birth. Thereafter the infants underwent BAL twice a week until 35 days of age or extubation, whichever occurred earlier. The local research ethics committee had approved the lavage procedure, and informed parental consent was obtained before the procedure was started. The collected BALF was placed on ice and centrifuged at 300 g at room temperature for 10 minutes within 10 minutes of collection. The supernatant was stored at −70°C until fibroblast assays were performed.

Fibroblast proliferation assays

Cell culture

Swiss 3T3 neonatal mouse lung fibroblasts (European Collection of Cell Cultures) were maintained in Dulbecco’s modification of Eagle’s medium containing penicillin (100 U/ml), streptomycin (100 µg/ml), amphotericin B (100 µg/ml), sodium pyruvate (1 mM), 1 × Glutamax II, and 1 × MEM non-essential amino acids (fibroblast growth medium (FGM)) supplemented with 10% fetal calf serum (FCS). All tissue culture reagents were supplied by BRL/Life Technologies (Paisley, Scotland, UK). Cell cultures were used between passages 6 and 10. The cells tested negative for mycoplasma contamination.

Microwell fibroblast proliferation cultures

Adherent cells were suspended with 0.02% trypsin/EDTA, and the reaction quenched with FGM/10% FCS. After centrifugation, the cells were resuspended in fresh FGM/10% FCS to give a final concentration of 5 × 10^3/200 µl. Then 200 µl of this cell suspension was introduced into 96 well plates. After overnight incubation at 37°C in 5% CO₂/95% air to permit cell adherence, the cells were quiesced in FGM/0.5% FCS for a further 24 hours. Thereafter, cells were incubated under the following experimental conditions: controls of 0.5%, 10% FCS, 20 ng/ml IGF-I or PDGF-BB; or BALF with or without growth factor and/or antibody (recombinant PDGF-BB, epidermal growth factor (EGF), and IGF-I and respective antibodies; R&D Systems Europe, Abingdon, Oxon, UK). All experiments were performed in triplicate.

Preliminary experiments showed that 48 hours of incubation was optimal to show fibroblast proliferation. Additional experiments showed that 1:10 to 1:20 dilution of BALF was optimal to show fibroblast proliferation (see below and fig 3). Thus, a 1:10 dilution was used for fibroblast proliferation assays in triplicate, and, because of limited volumes of BALF obtained from preterm infants, a 1:20 dilution was used in triplicate for antibody neutralising experiments.

Methylene blue assay

The proliferation of Swiss 3T3 cells was determined by a modified assay described by Oliver et al. After removal of the culture medium, the cells were fixed with 10% formaldehyde in phosphate buffered saline for one hour, and 100 µl 2% (w/v) fresh methylene blue in 0.1 M borate buffer (pH 8.5) was added. After elution of the dye from the cells with 1:1 (v/v) ethanol and 0.1 M HCl, the absorbance was measured at 650 nm with a microplate photometer (Dynatech Labs). Initial experiments validated the methylene blue assay with both cell counting and incorporation of [3H]thymidine. The results of the methylene blue assay were within 10% of the results obtained by cell counting and [3H]thymidine incorporation.

Antibody blocking experiments

Initial experiments using a dose range of 0–100 ng/ml recombinant growth factor (PDGF-BB and IGF-1) and the fibroblast proliferation assay described above showed that 20 ng/ml growth factor (both PDGF-BB and IGF-1) resulted in near maximum fibroblast proliferation. Additional experiments showed that 10× manufacturer’s specified neutralising dose of the respective antibody completely abolished the mitogenic activity of both growth factors, each used at 100 ng/ml. For experiments using BALF, the lavage fluid was incubated for two hours on ice with the antibody before addition to the fibroblasts. The experiments were terminated after 48 hours of incubation as described above by the addition of 10% formaldehyde in phosphate buffered saline. Cell proliferation was determined by using the methylene blue assay described above.

Data analysis

Results are given as median (range) for patient characteristics such as birth weight and gestation, and median (interquartile range) for the fibroblast proliferation assays. All experiments with the fibroblast proliferation assay were performed in triplicate. The mean A₆₅₀ of the triplicates is given for single experiments. When multiple plates were compared, the A₆₅₀

### Table 1 Patient characteristics

<table>
<thead>
<tr>
<th></th>
<th>CLD</th>
<th>RDS</th>
<th>Control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number</td>
<td>16</td>
<td>18</td>
<td>9</td>
</tr>
<tr>
<td>Male to female</td>
<td>9.7</td>
<td>11.7</td>
<td>6.3</td>
</tr>
<tr>
<td>Birth weight (g)</td>
<td>765 (630–1230)</td>
<td>1415 (430–4160)</td>
<td>2110 (900–3720)</td>
</tr>
<tr>
<td>Gestational age (weeks)</td>
<td>26.5 (23–29)</td>
<td>31 (23–39)</td>
<td>32 (26–41)</td>
</tr>
<tr>
<td>Antenatal dexamethasone</td>
<td>15 (94%)</td>
<td>8 (44%)</td>
<td>3 (33%)</td>
</tr>
<tr>
<td>Surfactant administration</td>
<td>15 (94%)</td>
<td>15 (83%)</td>
<td>1 (11%)</td>
</tr>
<tr>
<td>Day 1 maximum peak inspiratory pressure (cm H₂O)</td>
<td>17 (14–22)</td>
<td>19 (14–29)</td>
<td>15 (14–20)</td>
</tr>
<tr>
<td>Day 1 maximum FIO₂</td>
<td>0.40 (0.24–0.90)</td>
<td>0.41 (0.21–0.00)</td>
<td>0.24 (0.21–0.28)</td>
</tr>
<tr>
<td>Day 1 mean airway pressure (cm H₂O)</td>
<td>7.5 (5–21)</td>
<td>9.5 (4.4–29)</td>
<td>5.3 (4–7)</td>
</tr>
<tr>
<td>Day 1 maximum oxygenation index</td>
<td>5.3 (2.1–30.5)</td>
<td>5.9 (0.9–94.3)</td>
<td>1.8 (1.3–3.0)</td>
</tr>
</tbody>
</table>

Except where indicated otherwise, median values (range) are shown. CLD, Chronic lung disease of prematurity; RDS, respiratory distress syndrome.
Lung fibrosis in chronic lung disease

Figure 1. Relative fibroblast proliferation induced by bronchoalveolar lavage fluid obtained from infants who progressed to chronic lung disease of prematurity (CLD), who developed and recovered from neonatal respiratory disease (RDS), and control infants receiving mechanical ventilation for non-respiratory reasons. The proliferative activity is relative to the A_{650} value obtained for 10% fetal calf serum for each 96 well plate. *p < 0.01 for CLD v control and p < 0.05 for RDS v control.

Figure 2. Neutralising antibodies were used against platelet derived growth factor (PDGF-BB) and epidermal growth factor (EGF) to show whether bronchoalveolar lavage fluid (BALF) fibroblast activity was dependent on these two growth factors. Parallel experiments showed that BALF spiked with PDGF-BB did increase fibroblast proliferation, but this was decreased to baseline levels by prior addition of a neutralising antibody (ab) to PDGF-BB.

Figure 3. A typical experiment showing that bronchoalveolar lavage fluid (BALF) dilutions increased fibroblast proliferation and that spiking diluted BALF with insulin-like growth factor I (IGF-I) further increased this proliferation and that it was prevented by the prior addition of a neutralising antibody (ab) to IGF-I. Furthermore, this antibody also decreased the fibroblast activity of native BALF at 1:10 and 1:50 dilutions.

Table 1 shows patient characteristics including use of surfactant and antenatal corticosteroids. In particular, 15/16 (94%) in the CLD group, 15/18 (83%) in the RDS group, and 1/9 (11%) in the control group were treated with exogenous surfactant. Significant differences in gestation and birth weight were noted between infants in the CLD and RDS groups (p < 0.0005) and those in the CLD and control groups (p < 0.0001), but not between those in the RDS and control groups (birth weight, p = 0.13; gestation, p = 0.073). Significant differences were also found between the CLD and control groups for inspired oxygen (p < 0.001) and oxygenation index (p < 0.005), but not for peak inspiratory pressure in the first 24 hours of age, and between the RDS and control groups for inspired oxygen (p < 0.01) and oxygenation index (p < 0.01). The difference for peak inspiratory pressure in the first 24 hours of age between the RDS and control groups approached significance (p = 0.06).

RESULTS

Patient characteristics

A total of 207 BAL samples were obtained from 43 infants requiring mechanical ventilation. Thirty one BAL samples were excluded because of the small volume, repeated fungal infection with the fibroblast proliferation assay, sepsis, blood staining, or they were from control infants who required > 28% oxygen. Of the 43 infants studied, 34 developed respiratory failure due to RDS, and, of these, 16 (nine boys, seven girls) progressed to develop CLD, and 18 (11 boys, seven girls) recovered and were nursed in air by 28 days of age. Nine required mechanical ventilation for non-pulmonary reasons. The chest radiographs for the infants from the CLD and RDS groups on day 1 were very similar, showing features of the RDS, but by day 28 of age the chest radiographs for infants in the CLD group had abnormalities, with mild to moderate haziness being the most common finding. No infant had cystic changes on their radiograph. The chest radiographs for the control infants were reported to be normal.

Detection of fibroblast mitogenic growth factors in BALF

Using neutralising antibodies to PDGF-BB and EGF, we were unable to contribute any fibroblast proliferative activity in BALF to these two growth factors (fig 2). Parallel experiments “spiking” BALF with growth factor showed that the increased proliferation of fibroblasts with 20 ng/ml PDGF-BB was prevented by the prior addition of neutralising antibody to PDGF-BB. As expected, spiking with EGF did not increase fibroblast proliferation (data not shown).

Further experiments used neutralising antibodies to IGF-I to show whether this growth factor contributed to the mitogenic activity observed in BALF from ventilated infants. Fibroblast proliferative activity was shown in BALF by using dilutions of 1:10 to 1:50 (one individual experiment shown in fig 3). Spiking each diluted BALF with IGF-I increased the fibroblast mitogenic activity of the BALF, and this was...
pathological studies suggest that lung fibrosis is a prominent feature in the pathogenesis of CLD. Increased fibroblast numbers and increased deposition of components of the extracellular matrix have been observed in these studies. Fibroblast proliferation is not prominent in the development of CLD. In addition, antibody neutralising experiments showed that this proliferative activity was not due to PDGF-BB or EGF, and only 11–15% was attributable to IGF-I. Factors other than these growth factors appear to be responsible for promoting fibroblast proliferation in the lungs of newborn infants.

Pathological studies suggest that lung fibrosis is a prominent feature in the pathogenesis of CLD. Increased fibroblast numbers and increased deposition of components of the extracellular matrix have been observed in these studies. For lung fibrosis to occur, two events are likely: the extracellular matrix, which may be of greater importance than increased proliferation of fibroblasts.

As rapid lung growth is probably regulated by the growth factors we have studied, any additional growth factor activity resulting from repair/remodelling processes after acute lung injury may be difficult to identify. Another explanation for the discrepancy between the increased fibroblast numbers observed at autopsy of infants who have died from CLD when compared with control infants and our data showing minimal increased mitogenicity for fibroblasts may be the removal of fibroblasts from the lung by the process of apoptosis. Most studies have concentrated on the proliferation of fibroblasts, and yet few have studied the removal of this cell type from the lung. Clearly the balance between fibroblast proliferation and removal of these cells will affect whether or not lung fibrosis occurs.

Growth factors such as IGF-I and PDGF-BB are known mitogens for lung fibroblasts, but in BALF from those infants who did and did not develop CLD, we were unable to show any contribution by PDGF-BB, and only 10–15% activity was attributable to IGF-I. This is unlikely to be due to methodological problems because the increased proliferation caused by spiking the lavage fluid was prevented by the prior addition of the relevant neutralising antibody. The existence of inhibitory autoantibodies to PDGF-BB in BALF has been postulated. We would like to speculate that the lack of fibroblast proliferation caused by PDGF-BB was not due to the presence of inhibitors because the spiking of BALF resulted in fibroblast proliferation, as the presence of inhibitors would have prevented such an effect. Factors other than the growth factors that we have studied are known to promote proliferation of fibroblasts. Particular attention has recently focused on components of the clotting cascades. Thrombin has been shown to be a potent mitogen for fibroblasts and has been shown to be increased in respiratory diseases in adults.

We have used methylene blue to assess the proliferation of fibroblasts with BALF, FCS, and growth factors. It is a simple technique which gives repeatable results. We validated the technique using [3H]thymidine incorporation and found a good correlation between the two methods, as previously reported by Laurent’s group. However, we are aware that the staining is proportional to both cell volume and cell numbers, but the correlation between this method and incorporation of thymidine is reassuring. The antibody neutralising experiments were performed with spiking of the BALF with IGF-I and PDGF-BB as well as EGF (as a control). We found that addition of IGF-I and PDGF-BB increased fibroblast proliferation, but the simultaneous addition of neutralising antibody abrogated this increase thus validating the method.

In summary, we are not aware of any other study showing the proliferative activity of BALF obtained from infants who develop CLD. Fibroblast proliferation was remarkably similar in all infants studied, suggesting that fibroblast proliferation is not prominent in the development of CLD. In addition, we have only been able to account for 10–15% of the mitogenic activity for fibroblasts in lung lavage fluid from newborn infants. Factors other than IGF-I and
PDGF-BB are responsible for promoting fibroblast division in the newborn lung.

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REFERENCES