Metabolic and respiratory effects of theophylline in the preterm infant

Virgilio P Carnielli, Giovanna Verlato, Franca Benini, Katia Rossi, Monica Cavedagni, Marco Filippone, Eugenio Baraldi, Franco Zacchello

Abstract
Background—Methylxanthines are often administered to preterm infants for the treatment of apnoea.

Aims—To study the effects of theophylline on energy metabolism, physical activity, and lung mechanics in preterm infants.

Methods—Indirect calorimetry was performed for six hours before and after administration of a bolus of theophylline (5 mg/kg) in 18 preterm infants while physical activity was recorded with a video camera. Lung mechanics measurements were performed at baseline and 12 and 24 hours after theophylline treatment.

Results—Theophylline increased mean (SEM) energy expenditure by 15 (5) kJ/kg/day and augmented carbohydrate utilisation from 6.8 to 8.0 g/kg/day, but fat oxidation was unchanged. After theophylline treatment, preterm infants had faster respiration, lower transcutaneous CO2, and improved static respiratory compliance without increased physical activity.

Conclusions—A bolus of 5 mg/kg theophylline increased energy expenditure independently of physical activity, increased carbohydrate utilisation, and improved respiratory compliance. The increased energy expenditure could be detrimental to the growth of the preterm infant.

Subjects and methods

Subjects
Infants were recruited in the neonatal intensive care unit of the Department of Paediatrics of the University of Padova, Italy. Patients eligible for the study were free of congenital anomalies, with a gestational age between 31 and 34 weeks, an adequate weight for gestational age, in a stable clinical condition, and not requiring supplemental oxygen. All preterm infants were studied within 72 hours of birth. Theophylline was given for the prevention of apnoea of prematurity as aminophylline (Aminomal Malesci, Firenze, Italy) with a loading dose of 5 mg/kg intravenously over one to two minutes followed by a maintenance dose of 2 mg/kg intravenously every 12 hours. The infants were kept in servo controlled incubators set to a skin temperature of 36.2–36.5°C. The protocol was approved by the ethics committee of the University of Padova, and informed consent was obtained from the parents.

STUDY PROTOCOL
The study consisted of the continuous measurement of V02, carbon dioxide production (VCO2), physical activity, transcutaneous oxygen, and carbon dioxide tension for six hours before and six hours after the loading dose of theophylline. Respiratory mechanics was measured before theophylline administration and repeated 12 and 48 hours after. Vital signs were also simultaneously and continuously recorded. No changes in fluid therapy or nutrition were allowed during the 12 hour study involving indirect calorimetry, physiological measurements, and physical activity, so the infants received exactly the same nutrition before and after theophylline. All preterm infants during the study period received continuous intragastric feeding.
Table 1  Respiratory gas exchange data in 18 premature newborn infants before and after the administration of aminophylline

<table>
<thead>
<tr>
<th>Physical activity</th>
<th>Before</th>
<th>After</th>
<th>Mean difference</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>VO2 (ml/min/kg)</td>
<td>5.77</td>
<td>6.25</td>
<td>0.47 (0.18)</td>
<td>&lt;0.02</td>
</tr>
<tr>
<td>CO2 production (ml/min/kg)</td>
<td>5.08</td>
<td>5.66</td>
<td>0.57 (0.15)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Respiratory quotient</td>
<td>0.87</td>
<td>0.90</td>
<td>0.02 (0.01)</td>
<td>&lt;0.03</td>
</tr>
<tr>
<td>Energy expenditure (kJ/kg/day)</td>
<td>170</td>
<td>185</td>
<td>15 (5)</td>
<td>&lt;0.02</td>
</tr>
<tr>
<td>Transcutaneous PO2 (mm Hg)</td>
<td>67.8</td>
<td>67.2</td>
<td>−0.67 (2.35)</td>
<td>0.72</td>
</tr>
<tr>
<td>Transcutaneous PCO2 (mm Hg)</td>
<td>39.8</td>
<td>35.4</td>
<td>−4.48 (1.06)</td>
<td>&lt;0.02</td>
</tr>
</tbody>
</table>

Data are expressed as mean (SEM).

USA) pneumotachograph (Lilly type; linear to 10 litres/min), connected to a Validyne model DP 250-14 differential pressure transducer (linear to ±0.196 kPa). Airway pressure is measured during the occlusion through a port in the valve system, connected by stiff tubing to a model DP 250-32 differential pressure transducer (Validyne Engineering Corp, Northridge, California, USA). The dead space of the system (valve pneumotachograph) was 2.8 ml. No sedation was given to induce quiet sleep in the infants. Behavioral criteria were used to avoid the necessity of taking measurements during active sleep phases. Infants were studied inside the incubators in the supine posture, with the head in the neutral position and the mandible supported.

Table 2  Percentage of time before and after theophylline administration spent in the different physical activity phases

<table>
<thead>
<tr>
<th>Activity level</th>
<th>Before</th>
<th>After</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Level 0</td>
<td>74.8</td>
<td>75.0</td>
<td>0.75</td>
</tr>
<tr>
<td>Level 1</td>
<td>15.6</td>
<td>14.9</td>
<td>0.81</td>
</tr>
<tr>
<td>Level 2</td>
<td>6.4</td>
<td>6.5</td>
<td>0.70</td>
</tr>
<tr>
<td>Level 3</td>
<td>2.5</td>
<td>3.5</td>
<td>0.06</td>
</tr>
</tbody>
</table>

Data are expressed as means.

RESPIRATORY FUNCTION TESTING

Respiratory system compliance, respiratory system resistance, and expiratory time constant were measured immediately before and 12 and 48 hours after the first theophylline dose. The passive expiratory flow-volume technique (single breath occlusion), a modification of the method described in detail by LeSouef et al., was applied by using a computerised infant pulmonary function device (SensorMedics 2600, Yorba Linda, California, USA). With this method, a slide valve connected to an appropriately sized face mask (Vital Signs, Totowa, New Jersey, USA) briefly occludes the airway at end inspiration, producing relaxation of the respiratory muscles through the Hering-Breuer reflex, as previously described. The occlusion is then rapidly removed to allow passive expiration through an H Rudolph (Kansas City, Missouri,

Figure 1  Heart rate (histogram) and oxygen consumption (curves) before and after theophylline administration for each level of physical activity. Heart rate is significantly increased by theophylline (p < 0.001) and by the level of physical activity (p < 0.05). Oxygen consumption is also increased by theophylline (p < 0.05) and physical activity (p < 0.05).
Metabolic effects of theophylline

Table 3  Substrate utilisation before and after theophylline administration

<table>
<thead>
<tr>
<th></th>
<th>Before</th>
<th>After</th>
<th>Mean difference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Carbohydrate</td>
<td>6.8</td>
<td>8.0*</td>
<td>1.19 (0.18)</td>
</tr>
<tr>
<td>Fat oxidation</td>
<td>1.6</td>
<td>1.4†</td>
<td>−0.21 (0.16)</td>
</tr>
</tbody>
</table>

Data are expressed in g/kg/day as mean (SEM). *p < 0.05, †p > 0.05 compared with value before treatment.

Table 4  Effect of theophylline administration on respiratory function in 16 infants

<table>
<thead>
<tr>
<th></th>
<th>Before</th>
<th>After</th>
<th>Mean difference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rrs (kPa/l/s)</td>
<td>7.1</td>
<td>6.8†</td>
<td>−0.31 (0.09)</td>
</tr>
<tr>
<td>TV (m3/kg)</td>
<td>7.5</td>
<td>8.1†</td>
<td>0.58 (0.28)</td>
</tr>
<tr>
<td>Crs (ml/kPa/kgPa)</td>
<td>16.6</td>
<td>18.3*</td>
<td>1.73 (0.28)</td>
</tr>
</tbody>
</table>

Data are expressed as mean (SEM).

Discussion

We investigated in preterm infants whether theophylline increases energy expenditure, and whether this effect could be related to stimulation of physical activity. We performed indirect calorimetry continuously for 12 hours, and found that a single intravenous dose of theophylline significantly increased energy expenditure by 15 kJ/kg/day. This differs from previously published data, which showed either a much greater effect or no effect. VO2 rose by 20% after 48 hours of theophylline treatment in a study by Gerhardt et al and by 25% after 24–48 hours of low dose theophylline in a study by Milsap et al. Both studies measured short term respiratory gas exchange, and measurements were performed with the infants sleeping. If the daily energy expenditure is calculated from these changes in VO2, theophylline would increase the metabolic rate by about 41.8 kJ/kg/day. If these are realistic estimates, theophylline would have a considerable negative effect on the growth of the preterm infant, which could be detected clinically. In our study, theophylline increased energy expenditure by 15 kJ/kg/day. Such a difference could still result in a diminished weight gain, which may not be easily detected in the small preterm infant who quite often suffers from several conditions known to affect growth.

At variance with the above studies and our data, Field et al, who measured energy expenditure by the doubly labelled water technique, found no difference between preterm infants treated with theophylline and a control

(3) v 139 (3) beats/min, p < 0.001, before and after theophylline respectively. Heart rate and Vo2 were also significantly increased by theophylline (p < 0.001 and p < 0.05 respectively) and by physical activity levels (p < 0.03 and p < 0.05 respectively) (fig 1).

Mean (SEM) blood glucose concentration before theophylline administration was 2.60 (0.19) mmol/l, and it rose to 3.21 (0.26), 3.35 (0.33), and 3.13 (0.15) mmol/l at two, six, and 12 hours respectively after theophylline administration. These values were not significantly higher than the basal determination. Table 3 shows substrate utilisation. Before theophylline, mean (SEM) carbohydrate utilisation was 0.8 g/kg/day, and it rose to 8.0 g/kg/day (p < 0.05) with a mean difference of 1.19 (0.18). Mean fat oxidation was not altered by theophylline administration: 1.6 v 1.4 g/kg/day before and after respectively.

Lung mechanics was studied in 16 of the 18 infants, and table 4 gives the data obtained. Mean (SEM) static respiratory compliance showed a significant improvement from 16.6 to 18.3 ml/kPa/kg 12 hours after theophylline, with a mean (SEM) difference of 1.73 (0.28), and to 18.8 ml/kPa/kg at the 48 hour time point (p<0.05), with a mean difference of 2.23 (0.62). No significant changes were found in mean respiratory system resistance and expiratory time constant values after the treatment. The mean coefficient of variation was 10% for respiratory system compliance and 11% for respiratory system resistance measurements.
group. Several factors may explain this difference. Metabolic adaptation to theophylline may have occurred in the patients of Fjeld et al but not yet in ours. We studied the acute effects of a loading dose of theophylline soon after birth, whereas the infants in the study of Fjeld et al were on a complete oral feed at 2–5 weeks of age and were probably on a full course of theophylline when studied. Metabolic adaptation to xanthines has been extensively described, and it may result from the inhibition of adenosine receptor binding. Differences in the calculation of energy expenditure with the doubly labelled water technique may occur when RQs are changing. We found a significantly higher RQ after theophylline administration (table 1). Under these circumstances, if the doubly labelled water technique is used, an underestimation of energy expenditure can be expected in the group with the higher RQ—that is, the theophylline treated patients. We cannot prove, however, that the differences in RQ in our study would persist for long periods, as VO₂ and VCO₂ measurements were stopped six hours after theophylline administration. We believe that the increase in VCO₂ is a reflection of an increased metabolic rate, as VO₂ did not change from 30 minutes after theophylline administration until the end of the study (data not shown). A CO₂ wash out phenomenon resulting from an increased respiratory drive is likely to last for a few minutes only.

The increase in energy expenditure in our patients after theophylline treatment was associated with increased glucose utilisation. Methylxanthines are reported to increase glycogenolysis, and lipolysis is augmented in adults but not in preterm infants. Increased glycogenolysis may have enhanced carbohydrate utilisation in our patients. We found an increase in plasma glucose after theophylline treatment, as reported by others, but the difference did not reach statistical significance. We cannot provide any further biochemical data because blood sampling was limited as we feared that the pain induced by more frequent blood sampling would have influenced energy expenditure and physical activity.

Our second aim was to see if changes in energy expenditure could be related to the effect of theophylline on physical activity. At variance with animal studies, we did not find that a single dose of theophylline affects physical activity in preterm infants in the first 3 days of life, thus other factors may cause the augmented energy expenditure. Methylxanthines have an effect on the central nervous system and may also be responsible for changes in sleep stages. Some studies in preterm infants receiving theophylline report an increase in active sleep and wakefulness, and there is ample evidence on the effect of sleep stages on energy expenditure. We did not perform electroencephalograms, therefore we are not able to quantify clearly the amount of time spent in rapid eye movement sleep before and after theophylline treatment; however, wakefulness was not different. Apart from an increase in rapid eye movement sleep, other mechanisms may be responsible for the changes in energy expenditure, including a direct effect of theophylline at the cellular level and increased work by the respiratory muscles.

After theophylline administration, the patients had higher heart rates, faster respiratory rates, and a decrease in transcutaneous CO₂. These results confirm those from other studies. Both caffeine and theophylline produce an increase in minute ventilation, an increase in PaCO₂, and an increase in most indices of neural respiratory drive. Davi et al have investigated the effect of theophylline on breathing in newborn infants and found a decreased CO₂ threshold and an increased CO₂ sensitivity. Gerhardt et al observed a parallel shift in the slope of the CO₂ response curve after theophylline administration.

Lung mechanics measurements showed a significant improvement in respiratory compliance after treatment with theophylline. The underlying mechanism may be related to enhanced excursion of the diaphragm and inspiratory effort. Both effects may determine recruitment of lung units and improve mechanical effectiveness of the respiratory system. The potentiating effect of theophylline on lung mechanics in vivo shown in our study is consistent with previous findings in animal experiments, but differs from the study of Gerhardt et al, who failed to show any improvement in dynamic compliance after theophylline treatment in preterm infants. To evaluate dynamic compliance, it is necessary to monitor oesophageal pressure changes, which are difficult to measure in preterm infants because of chest wall distortion. Enhanced diaphragmatic contractility after theophylline administration worsens chest wall distortion, alters distribution of pleural pressure, and disguises changes in dynamic compliance. We used an occlusion technique which avoids the necessity to estimate pleural pressure. Baseline mean compliance values in this study (16.6 ml/kPa/kg) are closely similar to those found by Gappa et al (15.4 ml/kPa/kg) in healthy preterm infants using the same technique and are lower than normative values for full term infants.

In conclusion, we have found that theophylline given to slightly preterm infants improved lung compliance, increased energy expenditure by 15 kJ/kg/day, and modified substrate utilisation. The rise in energy expenditure was not associated with an increase in physical activity as found in animal studies. We believe that large controlled trials including a long term follow up are mandatory to verify the usefulness and side effects of aminophylline and to ascertain if theophylline treatment results in a clinically significant effect on the growth of the preterm infant. This, in our opinion, is justified by the extensive use of xanthines in the management of preterm infants.

We thank Giuseppe Torreisin and his staff for technical support during the validation of the indirect calorimeter.


