Effects of red cell transfusion on cardiac output and blood flow velocities in cerebral and gastrointestinal arteries in premature infants

Mathias Nelle, Christina Höcker, Eugen P Zilow, Otwin Linderkamp

Abstract
Anaemia may increase the risk of tissue hypoxia in preterm infants. The effect of transfusion on circulation was studied in 33 preterm infants with a mean (SD) gestational age of 29 (5) weeks (range 26–34), birth weight 1153 (390) g (range 520–1840), and postnatal age of 48 (21) days (range 19–100). Packed cell volume, blood viscosity (capillary viscometer), cardiac output, and cerebral blood flow velocities in the internal carotid artery, anterior cerebral artery, and coeliac trunk (Doppler ultrasound) were determined before and after transfusion of 10 ml/kg of packed red blood cells. Transfusion increased packed cell volume from a mean (SD) 0.27 (0.45) to 0.37 (0.48). Mean arterial blood pressure did not change while heart rate decreased significantly from 161 (14) l/min to 149 (12). Cardiac output decreased from 367 (93) ml/kg/min to 311 (74) due to decrease in stroke volume from 2.28 (0.57) ml/kg to 2.14 (0.46) and in heart rate. There was a significant increase in systemic red cell transport (cardiac output times packed cell volume) by 17%, systemic flow resistance (blood pressure to cardiac output ratio) by 23%, and blood viscosity by 33%. Vascular hindrance (flow resistance to blood viscosity ratio) did not change significantly, thereby suggesting that neither vasoconstriction nor vasodilatation occurred with transfusion. After transfusion blood flow velocities decreased significantly in the anterior cerebral artery by 23%, in the internal carotid artery by 8%, and in the coeliac trunk by 12%. Red cell transport estimated as products of blood flow velocities times packed cell volume increased significantly by 25% in the internal carotid artery and by 21% in the coeliac trunk. These results indicate that red cell transfusion improved systemic oxygen transport as well as oxygen transport in the internal carotid artery and coeliac trunk.

Patients and methods
PATIENTS
Thirty three preterm infants with a mean (SD) gestational age of 29 (5) weeks (range 26–34), birth weight 1153 (390) g (range 520–1840), postnatal age 48 (21) days (range 19–100), and actual body weight 1935 (432) g (range 1000–2700) were studied with the approval of the ethical committee of the University Hospital of Heidelberg and with informed consent of the parents. At the time of investigation, all infants were in a stable clinical condition without respiratory or cardiac problems, signs of sepsis, or renal disease. Arterial blood gas values were: pH 7.35–7.45, carbon dioxide tension 4.7–6.0 kPa, and oxygen tension 8.0–10.7 kPa. They were not ventilated and not treated with sedatives, diuretics, or vasoactive drugs. The infants received 10 ml/kg of pure red blood cells over six hours. The blood had been taken two to five days before transfusion from the donors. Cardiovascular measurements were done before and four hours after transfusion.

Blood samples for determination of venous packed cell volume and blood viscosity (2 ml of
antecubital blood) were collected before and four hours after transfusion. Excessive squeezing and prolonged tourniquet application were avoided. The blood samples were taken after the cardiovascular measurements had been done.

**HAEMATOLOGICAL METHODS**

Packed cell volume was measured in duplicate by the microhaematocrit method. The values were corrected for 2% of trapped plasma. Red blood cell count, haemoglobin concentration, and white cell count were determined using a cell counter (Contraves). Blood viscosity was measured within one hour after blood collection using a capillary viscometer.

**CIRCULATORY PARAMETERS**

The cardiovascular examinations were made at least an hour after previous feeding. Measurements were done when the infants were in a quiet sleep state. Quiet sleep was assumed when infants eyes were closed, no movements were observed apart from occasional stable reactions and respiration was regular. The infants were in the supine position. Blood flow velocities and cardiac output were measured using an Interspec XL pulsed Doppler ultrasound system (Interspec Inc) as previously described. The aortic valve annulus was visualised by M mode echocardiography using the parasternal long axis view, and the aortic diameter was measured by the leading edge method from the anterior aortic wall to the anterior boundary of the posterior aortic wall in late diastole over five consecutive cycles. Aortic cross sectional area was calculated in late diastole as $\pi \times r^2$, where $r$ represents the annular radius averaged over five consecutive cycles. From an apical four chamber view, the pulsed Doppler sample volume was placed at the level of the aortic valve annulus. Aortic velocity integrals were recorded with a mechanical 5-0 MHz transducer using the duplex mode in an attempt to obtain the fastest spectral envelopes. Doppler wave forms were analysed by the software of the ultrasound system for peak velocity, mean velocity, and average time velocity integral.

Cardiac output was calculated as the product of stroke volume and heart rate. Cardiac index was calculated as the ratio of cardiac output:actual body weight.

Cerebral blood flow velocities in the anterior cerebral artery and right and left internal carotid artery were measured using a 5-0 MHz pulsed Doppler transducer from a coronal scan via the anterior fontanelle. The arteries were identified by the duplex scan mode. There were no significant differences between the two internal carotid arteries. Therefore, only the results of the right carotid artery are shown.

The coeliac trunk was localised by ultrasound from a longitudinal abdominal section. Blood flow velocity in the coeliac trunk was determined close to the origin of the artery from the abdominal aorta. The direction of the blood flow in this region is directed towards the transducer. There was no significant angle of insonance between the Doppler beam and the coeliac artery. Angle corrections of velocities were made in all cases.

In all vessels the system software was used to calculate maximal systolic, maximal end diastolic, and mean average flow velocity. The average of five consecutive, homogeneous flow cycles was taken in all measurements after 60 constant flow velocity integrals were recorded. When beat-to-beat coefficient of variation was less than 5%, Doppler recordings were taken as stable.

In 12 infants cardiac output, cerebral and intestinal flow velocities were determined twice within one hour to estimate intraobserver variability of these parameters. Intraobserver variability was 9-8% for cardiac output, 6-5% for cerebral and 10-6% for intestinal blood flow velocities. To eliminate interobserver variability, all measurements were done by one person (CH).

**MISCELLANEOUS METHODS**

Mean arterial blood pressure was measured in the right and left upper arm using an oscillometric technique (Dinamap 847, Critikon). Some infants woke up during blood pressure measurements so that we cannot rule out that blood pressure increased in these infants. Systemic flow resistance (R) was calculated as mean pressure (P) to cardiac output ratio (Q = flow). According to the Hagen-Poiseuille law the flow resistance in a single vessel depends on blood viscosity ($\eta$), vessel length (L), and vessel radius (r): $R = \frac{P}{Q} = \frac{8\times L}{\pi \times r^4 \times \eta} = \frac{Z}{\eta}$

$Z = (8/\pi) \times (L/r^4)$ is the vascular hindrance and expresses the effect of the vessel geometry on the flow resistance. $Z$ can be calculated as resistance:viscosity ratio. $Z$ increases as a result of vasoconstriction and decreases with vasodilation. Although the Hagen-Poiseuille law is valid only for single circular vessels, $Z$ can also be used as an indicator of overall vasoconstriction when serial studies in the same persons are performed. As an index of red cell transport, the product of mean flow velocity, and peripheral venous packed cell volume was calculated. Systemic red cell transport was calculated as product of cardiac output and packed cell volume.

**STATISTICAL ANALYSIS**

A two tailed paired t test was done to test for differences in the measurements before and 4 hours after blood transfusion; $p<0.05$ was considered significant. Data are presented as mean (SD).

**Results**

Changes in haemodynamic and rheological variables after transfusion are shown in the
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**Discussion**

Anemia may predispose the preterm infant to tissue hypoxia because of reduced oxygen carrying capacity of the blood. The results of the present study show that in 3 to 14 week old preterm infants red cell transfusion decreased cardiac output due to decreases in heart rate and stroke volume suggesting that the heart rate before transfusion was increased in response to the anemia.\(^\text{16}\) As the rise in packed cell volume was more pronounced than the fall in cardiac output, systemic red cell transfusion (packed cell volume times cardiac output) increased significantly. Hudson et al found a decrease in cardiac output by 12% and a rise in red cell transport by 30% after transfusion of 20 ml/kg of red cells in preterm infants with 2 to 90 days of age.\(^\text{6}\) Alverson et al\(^\text{7}\) and Bifano et al\(^\text{8}\) found no significant change in cardiac output, but a rise in systemic red cell transport by about 25%.

Red cell transport in cerebral arteries showed a marked rise in the internal carotid artery (+25%) with transfusion, but no significant change in the anterior cerebral artery. Assuming that the blood flow velocities in cerebral arteries reflect the actual blood flows,\(^\text{17}\) this suggests that the overall oxygen transport to the brain was improved by transfusion, but with marked regional variations. In particular, the brain tissue supplied by the anterior cerebral artery does not appear to benefit from red cell transfusion. Rameakers et al studied the effect of transfusion on blood flow velocity in the pericallosal artery of preterm infants and found also no change in red cell transport.\(^\text{9}\)

Studies on the effect of transfusion on the blood flow velocity in the cerebral artery media of preterm infants do not appear to exist in the literature.

We observed a significant decrease (−12%) in blood flow velocity in the coeliac trunk and a rise in red cell transport (+21%). In newborn piglets, transfusion of 25 ml/kg of blood decreased cardiac output and gastrointestinal blood flow by 10%, whereas systemic and gastrointestinal red cell transport did not change.\(^\text{18}\)

As the arterial blood pressure did not change with transfusion, systemic vascular resistance (pressure to cardiac output ratio) increased after transfusion according to the decrease in cardiac output (table). Flow resistance increases with increasing vascular hinderance (that is, increasing vasoconstriction) and increasing blood viscosity.\(^\text{15}\)

In our study, red cell transfusion resulted in a marked rise of blood flow velocity whereas vascular hinderance did not change.\(^\text{18}\) We conclude that red cell transfusion did not change overall vessel geometry in the preterm infants.

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**Table**

<table>
<thead>
<tr>
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<th>Before transfusion</th>
<th>After transfusion</th>
<th>Difference</th>
<th>P Value</th>
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<tbody>
<tr>
<td>Packed cell volume (PCV)</td>
<td>0.27 (0.045)</td>
<td>0.37 (0.048)</td>
<td>0.1 (0.04)</td>
<td>0.001</td>
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<tr>
<td>Haemoglobin (g/dl)</td>
<td>0.09 (1.5)</td>
<td>0.22 (1.7)</td>
<td>0.34 (0.3)</td>
<td>0.001</td>
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<tr>
<td>Mean blood pressure (P, mm Hg)</td>
<td>9.6 (7.9)</td>
<td>10.1 (7.9)</td>
<td>0.7 (1.5)</td>
<td>NS</td>
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<tr>
<td>Heart rate (beats/min)</td>
<td>1.21 (123)</td>
<td>1.49 (123)</td>
<td>0.12 (3)</td>
<td>0.005</td>
</tr>
<tr>
<td>Cardiac output (CO, ml/kg/min)</td>
<td>376 (93)</td>
<td>311 (74)</td>
<td>66 (16)</td>
<td>0.005</td>
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<tr>
<td>Stroke volume (ml/kg)</td>
<td>2.28 (0.57)</td>
<td>2.14 (0.46)</td>
<td>0.14 (0.07)</td>
<td>0.05</td>
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<td>Blood viscosity (s, mPas)</td>
<td>0.13 (0.03)</td>
<td>0.24 (0.4)</td>
<td>0.09 (0.05)</td>
<td>0.001</td>
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<tr>
<td>Flow resistance (R=10^3×PICO)</td>
<td>160 (5)</td>
<td>196 (6)</td>
<td>36 (10)</td>
<td>0.005</td>
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<td>Vascular hinderance (R/h)</td>
<td>90 (27.2)</td>
<td>82.6 (26.4)</td>
<td>8 (5.3)</td>
<td>NS</td>
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<tr>
<td>Mean blood flow velocity (m/sec)</td>
<td>0.24 (0.06)</td>
<td>0.22 (0.05)</td>
<td>0.02 (0.01)</td>
<td>0.05</td>
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<td>Internal carotid artery</td>
<td>0.20 (0.06)</td>
<td>0.20 (0.06)</td>
<td>0.00 (0.01)</td>
<td>NS</td>
</tr>
<tr>
<td>Anterior cerebral artery</td>
<td>0.54 (1.3)</td>
<td>0.50 (0.9)</td>
<td>0.04 (0.02)</td>
<td>0.05</td>
</tr>
<tr>
<td>Coeliac trunk</td>
<td>0.54 (1.3)</td>
<td>0.50 (0.9)</td>
<td>0.04 (0.02)</td>
<td>0.05</td>
</tr>
<tr>
<td>Systemic red cell transport (CO×PCV)</td>
<td>991 (32.5)</td>
<td>115.9 (22.7)</td>
<td>16.8 (3.9)</td>
<td>0.005</td>
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<tr>
<td>Red cell transport</td>
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<tr>
<td>Internal carotid artery</td>
<td>6.5 (1.9)</td>
<td>8.1 (2.0)</td>
<td>1.6 (0.4)</td>
<td>0.005</td>
</tr>
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<td>Anterior cerebral artery</td>
<td>7.0 (2.2)</td>
<td>7.4 (2.7)</td>
<td>1.4 (0.7)</td>
<td>NS</td>
</tr>
<tr>
<td>Coeliac trunk</td>
<td>9.2 (3.4)</td>
<td>11.1 (3.0)</td>
<td>1.9 (0.4)</td>
<td>0.005</td>
</tr>
</tbody>
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